7-DEHYDROAGAPANTHAGENIN AND 8(14)-DEHYDROAGAPANTHAGENIN, TWO NEW SPIROSTAN SAPOGENINS FROM AGAPANTHUS AFRICANUS*

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Abstract—Besides sitosterol, yuccagenin (1a) and agapanthagenin (2a), the two new spirostan sapogenins 7-dehydroagapanthagenin (3a) and 8(14)-dehydroagapanthagenin (4a) have been isolated from the rhizomes of Agapanthus africanus and their structures determined.

INTRODUCTION

Agapanthus africanus Hoffmnsg., a plant of South African origin, was first studied by Takeda et al., who isolated yuccagenin (1a). Later, Stephen and Mathew, working with several unspecified species of Agapanthus, obtained 1a and the new spirostan sapogenin agapanthagenin (2a). The present paper reports our results of the unhydrolysed ethanolic extract of the rhizomes of A. africanus cultivated on the Canary Isles. In addition to sitosterol, 1a and 2a, we isolated the two new spirostan sapogenins 7-dehydroagapanthagenin (3a) and 8(14)-dehydroagapanthagenin (4a) whose structures were established as (25R)-spirost-7-en-2 α ,3 β ,5 α -triol and (25R)-spirost-8(14)-en-2 α ,3 β ,5 α -triol respectively. This is the first time that spirostan sapogenins with Δ 7 and Δ 8(14) are found in nature.

RESULTS AND DISCUSSION

- 7-Dehydroagapanthagenin (3a), C₂₇H₄₂O₅ (by MS), was eluted together with 2a, 8(14)-dehydroagapanthagenin (4a) and a spirostan sapogenin of still unknown structure, being
- * Part XXIII in the series "New Sources of Steriod Sapogenins". For Part XXII see González, A. G., Freire, R., Hernández, R., Salazar, J. A. and Suárez, E. (1973) Anal. Quím. 69, 1031.
- ¹ TAKEDA, K., OKANISHI, T. and SHIMAOKA, A. (1955) Ann. Rept. Shionogi Research Lab. 107; (1956) Chem. Abstr. **50,** 15916b.
- ² STEPHEN, T. (1956) J. Chem. Soc. 1167.
- ³ Mathew, G. E. A. and Stephen, T. (1957) J. Chem. Soc. 262.

separated by preparative column chromatography of the acetates on silica gel and silica gel-AgNO₃. 3a has three OH groups as inferred from its MS which shows the loss of 1, 2 and 3 H₂O molecules from the molecular ion. On mild acetylation, it gives the diacetate 3b, $C_{31}H_{46}O_7$, which has an OH function (IR: 3580 cm⁻¹), but its PMR spectrum does not present any signals assignable to protons geminal to it. Hence, one of the OH groups in 3a must be tertiary. The remaining two oxygens form part of a (20*S*, 22*R*, 25*R*)-spirostan ring; this is from the position and relative intensities of the bands at 982, 923, 900 and 865 cm⁻¹ in the IR spectrum of 3b, 4 and from the PMR signals (CDCl₃) at τ 6·55 (*m*, $W_{1/2}$ 12 Hz) and 8·39 (*s*, $W_{1/2}$ 5 Hz) characteristic of the 2H-C₂₆ and 2H-C₂₃ respectively.^{5.6}

Compound	Solvent	$H/C_{2,3}$	н С,	2HC ₂₀	$\operatorname{Me} \cdot C_{10}$	Mc-C ₁₃	Me-C ₂₀	Me C ₂₅	OAc
Agapanthagenin	CDCl ₄	4-77		6:55	8-92	9-23	9:05	~ 9-24	7-99
diacetate 3h		nt[25]		m[13]	5	S	d(6)		N
	C_0D_0	4.49		6:44	9-25	9-25	8:83	~ 9.30	8-24.
		m[36]		m[13]	¥	S	d(6)		8:27.
7-Dehydroagapanthagenin	$CDCI_k$	4.80	4.90	6.55	8-96	9:33	~ 9-02	9-21	7-98
diacetate 3b	•	m[24]	m[24]	m[12]	8	8		d(7)	S
	C_nD_n	4.50	5-13	6.42	9-22	9.34	8.83	~9.34	8-22
		m[36]	m[10]	m[12]	Δ	.5	d(6)		5
8(14)-Dehydroagapanthagenin	$CDCl_1$	4.78		6.55	9-02	9:02	~ 9:02	9-23	7.99
diacetate 4b		m[26]		m[13]	8	8		d(6)	S
	C_nD_n	4.50		6:43	9:00	9-24	8.85	~9-31	8-23
	- 0 0	mf 307		m[13]	\$	8	d(6)		5

TABLE 1. CHEMICAL SHIFTS IN THE PMR SPECTRA OF SPIROSTAN SAPOGENINS (T-scale, 60 MHz)

The molecular formulae for 3a and 3b together with the IR absorptions at 3025, 1670 and $840\,\mathrm{cm^{-1}}$ of the diacetate indicate the presence of a double bond which must be tertiary since in the PMR spectrum (C_6D_6) of 3b only one vinyl proton is observed (τ 5·13, m, $W_{1,2}$ 10 Hz). Taking into account that the chemical shifts and shapes of the signals for the protons geminal to the OAc groups in the PMR spectra of 3b (CDCl₃ and C_6D_6) coincide with those observed for agapanthagenin diacetate (2b) (see Table 1), one may deduce that 3a has a 2α , 3β , 5α -triol system.* On this basis the trisubstituted double bond can only be placed at C_7 as is inferred from PMR data: the theoretical values for the Me- C_{10} and Me- C_{13} of 3b (τ 8·93 and 9·35)† agree with the experimental ones (CDCl₃, Table 1).

Structure 3a was confirmed by the following reaction sequence: dehydration of 3b with $SOCl_2$ in pyridine at 0° gave a mixture of 7-dehydroyuccagenin acetate (5) and 4,7-dehydrogitogenin acetate (6), no appreciable changes in their ratio being observed on modifying the temperature or $SOCl_2$ concentration. The structure of 5, $C_{31}H_{44}O_6$, was established by spectral analysis: in the UV, typical absorptions were found at 261, 270, 281 and 292 nm, 8 and in the PMR spectrum (CDCl₃) the $H-C_{6,7}$ appear as an AB system; further-

^{*} Coupling constants J in parentheses, $W_{1/2}$ in brackets (both in Hz).

^{*} The strong deshielding observed for the H-C_{2,3} (\sim 0·36 ppm in CDCl₃ and C₆D₆) as compared with the chemical shifts of these protons in gitogenin acetate [(25R)-5 α -spirost-2 α ,3 β -diol acetate; τ 5·13 (CDCl₃) and 4·86 (C₆D₆)] is due to the presence of the 5 α -OH group and is valid for characterizing this system.

[†] Contributions for the double bonds are taken from Ref. 7.

⁴ Jones, R. N., Katzenellenbogen, E. and Dobriner, K. (1953) J. Am. Chem. Soc. 75, 158.

⁵ WILLIAMS, D. H. and BHACCA, N. S. (1965) Tetrahedron 21, 1641.

⁶ Callow, R. K., James, V. H. T., Kennard, O., Page, J. E., Paton, P. N. and Riva de Sanseverino, L. (1966) J. Chem. Soc. C, 288.

⁷ BHACCA, N. S. and WILLIAMS, D. H. (1964) *Application of NMR Spectroscopy in Organic Chemistry*, p. 19, Holden-Day, San Francisco.

⁸ ROSENKRANZ, G., ROMO, J. and BERLIN, J. (1951) J. Org. Chem. 16, 290.

more, the chemical shifts of the Me– C_{10} and Me– C_{13} (τ 8-94 and 9-28 in CDCl₃) coincide with the calculated ones (τ 8-94 and 9-27). 5 was also obtained by irradiating yuccagenin acetate (1b) in the presence of NBS,⁸ yielding 7 which without further purification was refluxed in o-xylene–collidine to give the expected 5 and 8. The latter compound shows UV absorptions at 233, 239 and 247 nm,⁸ and in the PMR spectrum the H– $C_{6,7}$ and H– C_4 appear as an ABX and AB system respectively.

Compound 6, $C_{31}H_{44}O_6$, has two trisubstituted double bonds evidenced by two one-proton signals at τ 4·76 (AB, $W_{1/2}$ 5 Hz, H–C₄) and 5·04 (m, $W_{1/2}$ 10 Hz, H–C₇) in the PMR spectrum (C_6D_6). Its structure was established by partial hydrogenation over 5% Pd–C in dry EtOAc, giving 8(14)- and 7-dehydrogitogenin acetates (9 and 10; both $C_{31}H_{46}O_6$);* the PMR spectrum of 10 shows the presence of a vinyl proton (τ 4·84, m, $W_{1/2}$ 10 Hz). The same two compounds were also formed by reducing 5 under identical conditions. The fact that under these conditions the double bond in 10 is not isomerized to $C_{8(14)}$ indicates that 9 is produced directly from 5 and 6. Transformation of 10 into 11 was achieved in

Chemical evidence for the α configuration of the OH at C_5 was obtained by treatment of 5 with 1·2 mol of *m*-chloroperbenzoic acid in dry benzene at 0–5°; the less hindered

^{*}About 40% of hydrogenolysis products were also formed, which is in accord with the existence of an allylic acetate in 6.

⁹ MANCERA, O., BARTON, D. H. R., ROSENKRANZ, G. and DJERASSI, C. (1952) J. Chem. Soc. 1021. PHYTO 13/3 G

double bond (Δ^5) is attacked and the epoxide enters from the α side giving 11. Subsequent reduction with LiAlH₄ and acetylation yielded 3b. Hence, the structure of 7-dehydroagapanthagenin (3a) is established as (25R)-spirost-7-en-2 α .3 β .5 α -triol.

8(14)-Dehydroagapanthagenin (4a). $C_{27}H_{42}O_5$ (by MS), has three OH groups, as shown by MS. One of them must be tertiary because 4a forms a diacetate (4b). $C_{31}H_{46}O_7$, with IR band at 3520 cm⁻¹ but whose PMR spectrum displays no signals for protons geminal to the OH group. Again the three OH functions form a $2\pi.3\beta.5\alpha$ -triol system (see Table 1) and the presence of a (20*S*, 22*R*, 25*R*)-spirostan ring is deduced from IR and PMR data as described for 3a. The molecular formulae of 4a and its diacetate indicate the existence of a double bond which must be tetrasubstituted because 4b shows no PMR signals attributable to vinyl protons. The only position for this double bond compatible with the PMR data is $C_{8(14)}$: the calculated values for the Me- C_{10} (τ 9·04) and Me- C_{13} (9·05) agree with the observed ones in CDCl₃ (Table 1). Structure 4a was confirmed by isomerizing the double bond in 3b to $C_{8(14)}$. As has been observed, ¹⁰ a Δ ⁷ is hindered by a 5 α -OH group; consequently, the isomerization conditions must be stronger than those required from the 5 α -H series. In fact, the transformation of 3b into 4b was only achieved in HOAc with 10% Pd-C under H_2 . Thus, 4a is assigned the structure of (25*R*)-spirost-8(14)-en-2 α ,3 β ,5 α -triol.

EXPERIMENTAL

M.ps, determined on a Kofler block, are uncorrected. If not otherwise stated compounds were recrystallized from MeOH. Optical rotations were measured in CHCl₃ and PMR spectra (60 MHz) with TMS as internal standard. The spray reagent for TLC was H₂SO₄-HOAc ·H₂O (1:20:4). Column and dry column chromatography was performed on silica gel 0·2-0·5 and 0·063-0·20 mm, respectively. Acetylations were realized with Ac₂O in pyridine and saponifications with 2°₀ KOH in MeOH, in both cases leaving the mixture at room temp. for 12 hr. Usual work-up was as follows: pour into H₂O, extract with organic solvent, wash with aq. HCl and H₃O, dry over Na₃SO₄ and evaporate in ractio.

Isolation of the sapogenins. The air-dried rhizomes of the plant (5 kg), collected in La Laguna (Tenerife) in August 1972, were finely chopped and extracted several times with EtOH in a Soxhlet. After filtering the combined extracts in cold, they were concentrated in racuo to call litre, diluted with an equal vol. of $\rm H_2O$ and defatted with $\rm C_6H_6$ in a liquid-liquid extractor. Then conc. HCl was added to the aq. ethanolic soln till it was 2 N. It was refluxed for 4 hr, poured into $\rm H_2O$, neutralized first with KOH and finally with NaHCO₃ and filtered. The precipitate was dissolved in EtOAc and washed several times with $\rm 10^{\circ}_{-0}$ aq. KOH and $\rm H_2O$. Evaporation of the solvent afforded the crude mixture of sapogenins (27·6 g) which on column chromatography with CHCl₃ and CHCl₃ EtOAc as cluants gave sitosterol (0·9 g), yuecagenin (1a; 3·2 g) and a mixture of four spirostan sapogenins. This was acetylated and chromatographed on a dry column ($\rm C_6H_6$ -EtOAc. 4:1), obtaining the pure $\rm 2x.3\beta$ -diacetates of 8(14)-dehydroagapanthagenin (4b: 0·23 g) and agapanthagenin (2b: 0·41 g). The remaining mixture of the acetates of 7-dehydroagapanthagenin (3b: 3·9 g) and a spirostan sapogenin currently under study (0·20 g) was separated by dry column chromatography on silica gel -20°₆ AgNO₃ cluting with $\rm C_6H_6$. EtOAc (9:1).

Yuccagenin Ia, m.p. 248–249". [z]_D = 120" (c 0·302). Acetate 1b, m.p. 179–181". [z]_D = 143" (c 0·214) (Found: C, 72·27; H, 8·74. Calc. for C_{31} H₄₆O₆: C, 72·34; H, 9·01°₆). v_{max}^{CS} : 3040, 2830 (Δ^5). 1745 (OAc). 980, 920, 900, 865 cm⁻¹ (spirostan ring). PMR (CDCl₃): τ 4·53 (1H, m, $W_{1/2}$ 1H Hz, H-C₆), 5·08 (2H, m, $W_{1/2}$ 30 Hz, H-C_{2/3}), 6·57 (2H, m, $W_{1/2}$ 13 Hz, 2H-C_{2/6}), 7·98 (6H, s, OAc), 8·88 (3H, s, Me-C_{1/6}), 9·03 (3H, d, d) 6 Hz, Me-C_{2/6}), 9·21 (3H, s, Me-C_{1/3}), \sim 9·21 (3H, d, Me-C_{2/5}). Hydrogenation of 1b (0·2/g) in HOAc (30 ml) over 10°₆ Pd-C (0·1/g) for 8 hr at room temp, and atm. pres. gave gitogenin acetate (0·15/g), identical with an authentic sample (m.m.p., TLC. IR spectra superimposable).

Agapanthagenin $2x.3\beta$ -diacetate **2**b. m.p. 298–300 . [z]_D = 100 (c0-676) (Found: C, 70-03; H. 8-90. Calc. for $C_{31}H_{48}O_7$; C, 69-89; H. 9-08%). v_{max}^{KBr} ; 3490 (OH), 1740 (OAc), 980, 920, 900, 865 cm⁻¹ (spirostan ring). PMR: Table 1.

Conversion of 1b in 2b. 1b (0·46 g) in CHCl₃ (10 ml) was treated with m-chloroperbenzoic acid (0·52 g) in CHCl₃ (25 ml) at 0°. After 4 hr the soln was poured into H_2O , washed with aq. KOH and H_2O and the solvent evaporated in vacuo. The resulting crude 5z, 6z-epoxy derivative (0·41 g) was dissolved in El_2O (17 ml) and poured dropwise into a stirred suspension of LiAlH₄ (0·25 g) in El_2O (13 ml). After 2 hr at reflux the excess LiAlH₄ was destroyed with some drops of H_2O . Usual work-up gave a residue which was acetylated and purified by dry column

¹⁰ Bladon, P., Clayton, R. B., Greenhalgh, C. W., Henbest, H. B., Jones, E. R. H., Lovell, B. J., Silverstone, G., Wood, G. W. and Woods, F. (1952) J. Chem. Soc. 4883.

chromatography (C₆H₆-EtOAc, 4:1), yielding 2b (0·21 g), identical with the natural product (m.m.p., TLC, IR spectra superimposable).

7-Dehydroagapanthagenin 2α,3β-diacetate 3b, m.p. 223–225°, [α]_D -88° (c0·212) (Found: C, 69·91; H, 8·65. $C_{31}H_{46}O_7$ requires: C, 70·16; H, 8·74%), $v_{max}^{CS_3}$: 3580 (OH), 3025, 1670, 840 (Δ^7), 1745 (OAc), 982, 923, 900, 865 cm⁻¹ (spirostan ring). PMR: Table 1. Saponification gave 3a, m.p. 276–278° (Me₂CO); m/e (%) 446 (M⁺, 0.5), 428 (74), 410 (2), 392 (0.6).

8(14)-Dehydroagapanthagenin $2\alpha,3\beta$ -diacetate 4b, m.p. $272-273^{\circ}, [\alpha]_D - 64^{\circ}$ (c 0·208) (Found: C, 69·95; H, 8·52. C₃₁H₄₆O₇ requires: C, 70·16; H, 8·74%), $\nu_{\rm max}^{\rm KB}$: 3520 (OH), 1740 (OAc), 980, 920, 900, 865 cm⁻¹ (spirostan ring). PMR: Table 1. Saponification gave 4a, m.p. 264–267°; m/e (%) 446 (M⁺, 0·3), 428 (100), 410 (3), 392 (1).

7-Dehydroyuccagenin and 4,7-dehydrogitogenin acetates 5 and 6 from 3b. 3b (0·4 g) in dry pyridine (10 ml) was treated with SOCl₂ (0·3 ml) at 0° for 1·5 hr. Dry column chromatography of the product (0·35 g) on silica gel-20% $AgNO_3$ (C₆H₆-EtOAc, 19:1) gave 5 (0:11 g) and 6 (0:14 g).

5, m.p. 178–182, $[\alpha]_D = 147^\circ$ (c 0·200) (Found: C, 72·67; H, 8·84. $C_{31}H_{44}O_6$ requires: C, 72·63; H, 8·65%). $v_{max}^{CS_2}$: 3040, 2800, 1655, 840 ($\Delta^{5.7}$), 1745 (Δ Ac), 985, 925, 900, 865 cm⁻¹ (spirostan ring). $\lambda_{max}^{E:OH}$: 261, 270, 281, 292 nm ($\Delta^{5.7}$). PMR (CDCl₃): τ 4·34, 4·44, 4·58, 4·68 (2H, AB, H–C_{6.7}), 5·07 (2H, m, W_{1.2} 27 Hz, $H-C_{2.3}$), 6·57 (2H, m, $W_{1,2}$ 12 Hz, 2H- C_{26}), 7·97 (6H, s, OAc), 8·94 (3H, s, Me- C_{10}), ~9·01 (3H, d, Me- C_{20}), ~9.23 (3H, d, Me-C₂₅), 9.28 (3H, s, Me-C₁₃); (C₆D₆): τ 4.7 (4H, m, W _{1/2} 36 Hz, H-C_{2,3,6,7}), 6.43 (2H, m, W _{1/2} 1.25) 12 Hz, 2H-C₂₀), 8·20 (6H, s, OAc), 8·82 (3H, d, J 6 Hz, Me-C₂₀), 9·08 (3H, s, Me-C₁₀), 9·28 (3H, s, Me-C₁₃), ~ 9.28 (3H, d, Me-C₂₅).

6, m.p. 200–202°, $[\alpha]_D - 119^\circ$ (c 0·382) (Found: C, 72·43; H, 8·53. $C_{31}H_{44}O_6$ requires: C, 72·63; H, 8·65%). $\frac{c_{33}^{CS}}{max}$: 3030, 2830, 1670, 840 ($\Delta^{4.7}$), 1745 (OAc), 981, 922, 900, 865 cm⁻¹ (spirostan ring). PMR (CDCl₃): τ 4·52, 4-64 (1H, mm, $W_{1/2}$ 5 Hz, H-C₃), 4-83 (3H, m, $W_{1/2}$ 12 Hz, H-C_{2,4,7}), 6-56 (2H, m, $W_{1/2}$ 13 Hz, 2H-C₂₆), 7-94 (6H, s, OAc), 8·80 (3H, s, Me-C₁₀), 9·00 (3H, d, J 6 Hz, Me-C₂₀), 9·21 (3H, d, J 6 Hz, Me-C₂₅), 9·32 (3H, s, Me-C₁₀) C_{13}); (C_6H_6) : τ 4·17, 4·31 (1H, mm, $W_{1/2}$ 5 Hz, H-C₃), 4·56 (1H, m, $W_{1/2}$ ~24 Hz, H-C₂), 4·76 (1H, AB, $W_{1/2}$ 5 Hz, H-C₄), 5·04 (1H, m, $W_{1/2}$ 10 Hz, H-C₇), 5·36 (1H, m, $W_{1/2}$ 22 Hz, H-C₁₆), 6·42 (2H, m, $W_{1/2}$ 12 Hz, 2H-C₂₆), 2·104 (1H, m, $W_{1/2}$ 10 Hz, H-C₇), 5·36 (1H, m, $W_{1/2}$ 20 Hz, H-C₁₆), 6·42 (2H, m, $W_{1/2}$ 12 Hz, 2H-C₂₆), 2·104 (1H, m, $W_{1/2}$ 10 Hz, H-C₁₆), 6·42 (2H, m, $W_{1/2}$ 10 Hz, 2H-C₂₆), 2·104 (1H, m, $W_{1/2}$ 10 Hz, H-C₁₆), 6·42 (2H, m, $W_{1/2}$ 10 Hz, 2H-C₂₆), 2·104 (1H, m, $W_{1/2}$ 10 Hz, H-C₁₆), 6·42 (2H, m, $W_{1/2}$ 10 Hz, 2H-C₂₆), 2·104 (1H, m, $W_{1/2}$ 10 Hz, H-C₁₇), 3·104 (1H, m, $W_{1/2}$ 10 Hz, H-C₁₆), 6·42 (2H, $W_{$ 8·21 (6H, s, OAc), 8·81 (3H, d, Me- C_{20}), 8·98 (3H, s, Me- C_{10}), 9·33 (3H, s, Me- C_{13}), \sim 9·33 (3H, d, Me- C_{25}).

7-Dehydroyuccagenin and 4.6-dehydrogitogenin acetates (5 and 8) from 1b. 1b (0·15 g) in dry CCl₄ (10 ml) was refluxed with NBS (0.075 g) for 5 min, irradiating with a W lamp (60 W). The soln was poured into H₂O, CHCl₃ extracted and the organic layer washed with aq. NaHCO₃ and H₂O. Evaporation in vacuo gave the 7α-bromo derivative 7 (0.15 g) which without further purification was dissolved in o-xylene (5 ml) and collidine (0.5 ml) and refluxed for 30 min. Dry column chromatography of the product on silica gel ~20% AgNO₃ (C₆H₆-EtOAc, 19:1) yielded pure 5 (61 mg) and 8 (42 mg).

5, m.p. 179–183°, $[\alpha]_D = 142^\circ$ (c.0-184) (Found: C, 72-37; H, 8-81. $C_{31}H_{44}O_6$ requires: C, 72-63; H, 8-65%).

Identical with compound 5 obtained from 3b (m.m.p., TLC, UV, IR and PMR spectra superimposable). 8. m.p. 204–207°, [α]_D -225° (c 0·276) (Found: C, 72·85; H, 8·68. $C_{31}H_{44}O_6$ requires: C, 72·63; H, 8·65%). v_{max}^{CS} : 3025, 1655, 855 ($\Delta^{4.6}$), 1745 (OAc), 982, 920, 900, 865 cm⁻¹ (spirostan ring). λ_{max}^{EOH} : 233, 239, 247 nm $(\Delta^{4.6})$. PMR(CDCl₃): τ 3·94, 4·11 (1H, ABX, J_{AX} 4 Hz, H-C₇), 4·22, 4·38 (1H, ABX, $W_{1/2}$ 4 Hz, H-C₆), 4·52 (1H, AB, $W_{1/2}$ 6 Hz, H-C₄), 4-76 (2H, m, $W_{1/2}$ 20 Hz, H-C_{2,3}), 6-56 (2H, m, $W_{1/2}$ 12 Hz, 2H-C₂₆), 7-96 (6H, s, OAc), 8.88 (3H, s, Me-C₁₀), 9.03 (3H, d, J 6 Hz, Me-C₂₀), 9.16 (3H, s, Me-C₁₃), \sim 9.21 (3H, d, Me-C₂₅).

8(14)- and 7-dehydrogitogenin acetates (9 and 10) from 6. 6 (90 mg) in EtOAc (50 ml) was hydrogenated for 3 hr over 5% Pd-C (47 mg) at room temp, and atm. pres. Separation of the residue by dry column chromatography (silica gel-20% AgNO₃; C_6H_6) gave hydrogenolysis products (34 mg) and pure 9 (11 mg) and 10 (17 mg).

9, m.p. 214–216°, [α]_D – 56° (c 0·132) (Found: C, 72·20; H, 9·11. $C_{31}H_{46}O_6$ requires: C, 72·34; H, 9·01%). $v_{\text{max}}^{\text{CS}_2}$. 1740 (OAc), 982, 925, 900, 865 cm⁻¹ (spirostan ring). PMR (CDCl₃): τ 5-08 (2H, m, $W_{1/2}$ 24 Hz, H-C_{2,3}), 6-56 $(2H, m, W_{1/2}, 13 \text{ Hz}, 2H-C_{26}), 8.00 \text{ (6H, s, OAc)}, 9.04 \text{ (3H, s, Me-C_{10})}, \sim 9.04 \text{ (3H, d, Me-C_{20})}, 9.20 \text{ (3H, s, Me-C_{20})}$ C_{13}), ~9.21 (3H, d, Me- C_{25}).

10, m.p. 221–225°, $[\alpha]_D$ –103° (c 0·216) (Found: C, 72·08; H, 8·89. $C_{31}H_{46}O_6$ requires: C, 72·34; H, 9·01%). $v_{\text{max}}^{\text{CS}_2}$: 3030, 1710, 845 (Δ^7), 1745 (OAc), 982, 925, 900, 865 cm⁻¹ (spirostan ring). PMR (CDCl₃): τ 4·84 (1H, m, $W_{1/2}$ 10 Hz, H-C₇), 5·11 (2H, m, H-C_{2,3}), 6·56 (2H, m, $W_{1/2}$ 12 Hz, 2H-C₂₆), 8·00 (6H, s, OAc), ~9·03 (3H, d, $Me-C_{20}$), 9.08 (3H, s, $Me-C_{10}$), 9.22 (3H, d, J 6 Hz, $Me-C_{25}$), 9.36 (3H, s, $Me-C_{13}$).

9 and 10 from 5. 5 (75 mg) in EtOAc (25 ml) was hydrogenated for 3 hr over 5% Pd-C (40 mg) as described for 6. Dry column chromatography (silica gel-20% AgNO₃; C₆H₆) of the residue gave 9 (18 mg), m.p. 217-218°, and 10 (47 mg), m.p. 221-225°, identical with the compounds obtained above (m.m.ps, TLC, IR, PMR spectra superimposable).

9 from 10. A soln of 10 (30 mg) in HOAc (30 ml) was stirred for 20 hr with 10% Pd-C (20 mg) under H₂ at room temp, and atm. pres. Purification of the residue by preparative TLC (silica gel PF₂₅₄₊₃₆₆, thickness 0.5 mm; C₆H₆-EtOAc, 19:1) gave 9 (20 mg) which proved to be identical with the compound obtained from 5 and 6 (m.m.p., TLC. IR, PMR spectra superimposable).

3b from 5. A soln of 5 (51 mg) in dry C₆H₆ (14 ml) was treated with m-chloroperbenzoic acid (22 mg) in C₆H₆ (4 ml) at 0-5° for 4 hr. The resulting 5α,6α-epoxide 11 was recovered as described for the conversion of 1b in 2b, dissolved in Et₂O (14 ml) and reduced with LiAlH₄ (40 mg) in Et₂O (10 ml) at reflux for 2 hr. After usual work-up the residue was acetylated and purified by dry column chromatography (C₆H₆-EtOAc, 17:3) obtaining 3b (23 mg), m.p. $222-224^{\circ}$, $[\alpha]_D = 88^{\circ}$ (c 0·230), identical with the acetate of the natural product (m.m.p., TLC, IR, PMR spectra superimposable).

4b from 3b. 3b (100 mg) in HOAc (30 ml) was isomerized over 10% Pd–C (200 mg) for 24 hr as described for 9. Dry column chromatography (C_6H_8 . EtOAc. 17:3) of the residue gave starting material (10 mg) and 4b (30 mg). m.p. 272–273°, [α]_D = 70° (c 0·112) (Found: C. 70·28; H. 8·72. $C_{31}H_{46}O_7$ requires: C. 70·16; H. 8·74%). It showed to be identical with the acetate of the natural product (m.m.p., TLC, IR, PMR spectra superimposable).

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